

Tensile and Thermal Properties of Crosslinked Chitosan/Empty Fruit Bunch Biofilms by Phthalic Anhydride

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Abstract: *Chitosan (CS) has high potential applications in packaging, agriculture and food. However, the cost associated with CS application is high. Therefore, the addition of agriculture waste such as empty fruit bunch (EFB) in CS has been studied in order to reduce the cost of CS. The CS/EFB biofilms are prepared through solvent casting method. This paper studies the effect of phthalic anhydride (PA) crosslinking agent on tensile, morphological and thermal properties of CS/EFB biofilms. The results indicated that the tensile strength and elongation at break of uncrosslinked CS/EFB biofilms decreases. However, the modulus of elasticity increases with the increasing EFB content. In addition, the glass transition temperature (T_g) of CS/EFB biofilms increased with EFB content. The crosslinked CS/EFB biofilm with PA showed higher tensile properties (i.e., tensile strength, elongation at break and modulus of elasticity) in comparison with uncrosslinked biofilms. The T_g of crosslinked CS/EFB biofilms, meanwhile, was higher than the uncrosslinked biofilms due to formation of amide linkages. The formation of amide bonds between CS and PA was analysed using the Fourier transform infrared (FTIR) spectroscopy. The better filler dispersion and filler-matrix adhesion were proven by scanning electron microscopy (SEM). It was further observed that the percentage gel fraction of both uncrosslinked and crosslinked CS/EFB biofilms increased with increasing of EFB content. On the other hand, the crosslinked CS/EFB biofilms exhibited higher gel fraction than uncrosslinked biofilms due to the formation of amide crosslinkages in crosslinked biofilms.*

Keywords: Chitosan, empty fruit bunch, crosslinking, phthalic anhydride, biofilms

1. INTRODUCTION

In recent years, the environmental impact caused by non-biodegradable plastic wastes has become an increasing concern globally. A lot of research has been done to develop renewable and environmental friendly bio-based polymeric materials as potential replacement for non-biodegradable plastic materials.¹ Normally, these materials are made from renewable or natural resources such as wheat proteins, corn zein, gelatin, whey proteins, cellulose derivatives and chitosan (CS) are which non-toxic, biodegradable and edible.² Moreover, these biopolymers generally exhibit high biocompatibility, biodegradability and renewability.³

CS is a natural polymer which can be obtained through deacetylation of chitin. CS is a renewable source and is abundantly available.^{4,5} The CS films are generally non-toxic, biocompatible and possess antimicrobial properties.⁶ Furthermore, CS films are tough, long lasting, flexible, and show good tear resistance.⁷ Because of these properties, CS is suitable material for designing packaging application.^{8,9}

Natural fibres today are considered as a potentially genuine alternative to glass fibres as reinforcement in composite materials. Advantages of natural fibres over glass fibres include low cost, low density, resistance to breakage during processing, high strength-to-weight ratio, low energy content and recyclability.^{10,11} Oil palm empty fruit bunch fibres (EFBs) are available in abundance, in addition to being renewable, nontoxic, and less costly.¹² In Malaysia, the palm oil industry is a major producer and exporter.¹³ For instance, in 2009, 85.71 million tons of fresh fruit bunches (FFBs) were produced, and an estimated amount of 6.76 million tons of dried EFBs were generated from oil palm mills.¹⁴ Therefore, the EFBs have a potential to yield up to 73% of fibres to be used as natural fillers in biocomposites.^{15,16}

Due to poor tensile properties of uncrosslinked CS/EFB biofilms, the crosslinking agent was used in order to improve the properties of biofilms. It has been reported in our previous study that the utilisation of different crosslinking agent¹⁷⁻¹⁹ and addition of modifier⁴ in CS biocomposite films improved the tensile and thermal properties of biocomposite films.

To date, the study of EFB filled CS biofilms is not yet reported. Therefore, this paper focuses on the preparation of EFB/CS biofilms. In this study, the phthalic anhydride was used as crosslinking agent to enhance the tensile, thermal and morphological properties of EFB/CS biofilms.

2. EXPERIMENTAL

2.1 Materials

CS was purchased from Hunza Nutraceutical Sdn. Bhd. with average particle size and degree of deacetylation of 80 μm and 90%, respectively. EFB was collected from Malaysia Palm Oil Board, Bangi, Selangor, Malaysia. The EFB was cleaned, crushed and ground into powder form with the particle size of 38 μm . Acetic acid and phthalic anhydride (PA) were supplied by Sigma-Aldrich, Penang, Malaysia.

2.2 Preparation of Uncrosslinked and Crosslinked CS/EFB Biofilms

First, the CS powder was dispersed into acetic acid (1 v/v %) and then stirred for 30 min. The EFB powder was then added into CS solution (1.5 w/v %) and stirred for 15 min. For crosslinked CS/EFB biocomposite films with PA, the PA powder was first dissolved in ethanol to produce 1 w/v % of PA solution. Then, 1% of PA solution was mixed with CS solution and stirred for 30 min and then the EFB was added. These uncrosslinked and crosslinked CS/EFB solutions were poured into plastic acrylic mold and dried in an oven at temperature 40°C for 24 h. Table 1 listed the formulations of uncrosslinked and crosslinked CS/EFB biofilms with PA.

Table 1: Formulations of uncrosslinked and crosslinked CS/EFB biofilms with PA.

Materials	Uncrosslinked CS/EFB biofilms	Crosslinked CS/EFB biofilms
CS (wt%)	100, 90, 80, 70, 60	100, 90, 80, 70, 60
EFB (wt%)	0, 10, 20, 30, 40	10, 20, 30, 40
PA (%)	–	1

2.3 Tensile Properties

The tensile test of CS/EFB biofilms was carried out using an Instron Universal Testing machine, Model 5569, following the ASTM D 882. The specimens were cut into rectangular shape in the size of 100 \times 15 mm. Besides, the testing was performed at 25 \pm 3°C with the cross-head speed of 15 mm min⁻¹. Ten specimens were tested for each sample and the average were calculated.

2.4 Fourier Transform Infrared (FTIR) Analysis

The FTIR analysis was done using Perkin-Elmer, Model L1280044 instrument. The attenuated total reflectance (ATR) method was used with a spectrum

resolution of 4 cm^{-1} . The 16 scans in the wavelength range from 4000 to 600 cm^{-1} were recorded.

2.5 Gel Fraction

The gel fraction of each biocomposite film was determined according to the method by Ramaprasad et al.²⁰ The specimens were swell in 1% of acetic acid for 24 h. After that, the crosslinked portion (gel) was remains insoluble. Then, the gel was filtered and dried in oven at 50°C for 24 hours. The percentage of gel fraction can be calculated using equation below:

$$\%GF = \frac{w}{w_s} \times 100 \quad (1)$$

where,

$\%GF$ = percentage of gel fraction,

w = weight of gel, and

w_s = weight of specimen before swelling

2.6 Differential Scanning Calorimetry (DSC)

The DSC was performed using a DSC Q 10 research instrument. The CS/EFB biofilms were cut into small pieces and weight in range of $7 \pm 2\text{ mg}$. The two runs of heating and cooling scan was employed, which is first heating scanned from 30°C to 200°C and cooling scanned from 200°C to 30°C ; second heating from 30°C to 200°C , with a heating rate of $10^\circ\text{C min}^{-1}$ in nitrogen environment. The glass transition temperature (T_g) of CS/EFB biofilms were determined from DSC data.

2.7 Scanning Electron Microscopy (SEM)

The tensile fracture surface of CS/EFB biofilms was evaluated by using a scanning electron microscope (SEM) Model JSM-6460LA. The fracture ends of specimens were sputter coated with a thin layer of palladium to avoid electrostatic charging during examination.

3. RESULTS AND DISCUSSION

3.1 Tensile Properties

Figure 1 exhibits the tensile strength of uncrosslinked and crosslinked CS/EFB biofilms with PA at EFB content from 10 to 40 wt%. The tensile strength of uncrosslinked CS/EFB biofilms decreased with increasing of EFB content due to poor filler-matrix interaction and poor dispersion of EFB filler. Besides, it was found that at similar EFB content, the tensile strength of crosslinked CS/EFB biofilms with PA is higher than uncrosslinked biofilms. Moreover, at 10 wt% of EFB content of crosslinked CS/EFB biofilms showed higher tensile strength than neat CS film. This is due to the formation of amide linkages between PA and CS in crosslinked biofilm.

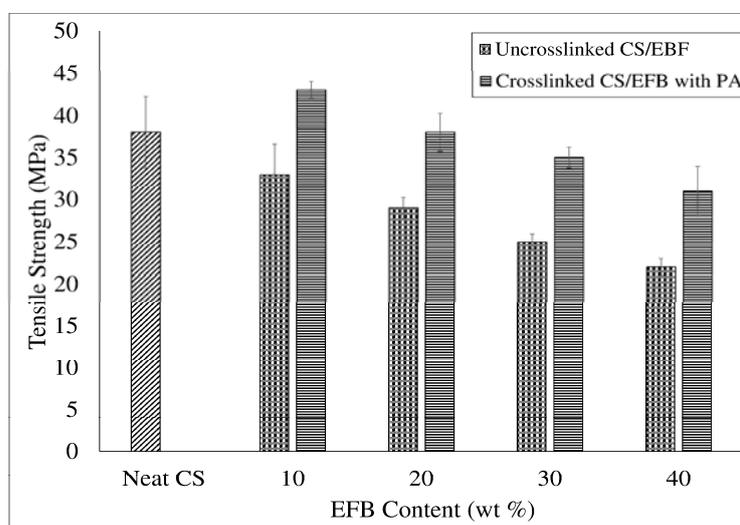


Figure 1: Tensile strength of uncrosslinked and crosslinked CS/EFB biofilms with PA.

The formation of crosslinkages in crosslinked biofilms cause the biofilms more shrinks than uncrosslinked biofilms, providing a compressive force as the CS matrix clamp down around the EFB fillers and consequently enhanced the filler-matrix interaction. These crosslinkages improved the filler-matrix interfacial interaction, and thus improved the tensile strength of CS/EFB biofilms. Besides, the chemical modification of CS/EFB biofilms with PA has also enhanced filler dispersion in CS/EFB biofilms. The average tensile strength of crosslinked CS/EFB biofilms with PA is 34.86% higher than uncrosslinked biofilms. Similar results have been reported by Yeng et al.,¹⁷ who studied the effect of corn cob content and glutaraldehyde as a crosslinking agent on CS/corn cob biocomposite films.

Figure 2 shows the influence of EFB content on the elongation at break of CS/EFB biofilms. The elongation at break of CS/EFB decreased with increasing EFB content due to the presence of natural filler, which reduced the chain mobility of CS polymer and increased the rigidity of CS/EFB biocomposite films.^{4,5,19} As the EFB content increased, the weak of interfacial region between filler surface and CS matrix were formed. These occurred because cracks travel more easily through the weaker interfacial region, resulting in lower elongation with the increasing EFB content. Interestingly, the elongation at break of crosslinked biofilms was higher than the uncrosslinked biofilms. This is attributed to the fact that chemical modification of biofilms with PA improved filler dispersion and thus increased the plasticisation effect of the crosslinked CS/EFB biocomposite films. Accordingly, Dearmitt²¹ noted that the better dispersant help to prevent agglomeration, decrease the effective particle size and thus help maintaining good elongation at break.

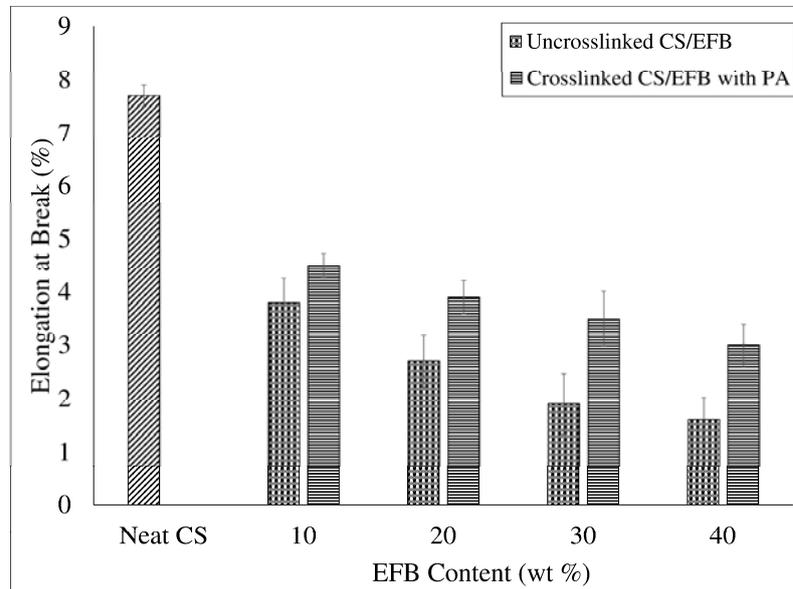


Figure 2: Elongation at break of uncrosslinked and crosslinked CS/EFB biofilms with PA.

Increase in EFB content resulted in a higher modulus of elasticity of CS/EFB biofilms, as shown in Figure 3. This result indicates that the introduction of EFB fillers decreased the flexibility of CS chain, resulting in higher stiffness of biocomposite films. Generally, the modulus of the filler is higher than the polymer matrix; thus, one of the functions of filler is to enhance the modulus of biocomposites.²² Consequently, the increment on elasticity modulus is a usually consistent behaviour in filler particle studies. As can be seen, at a similar EFB content, crosslinked CS/EFB biofilms exhibited higher modulus of elasticity than

uncrosslinked biofilms due to formation of amide crosslinkages in biocomposite films. The crosslinked CS/EFB biofilms with PA showed around 9.92% improvement in modulus of elasticity as compared to uncrosslinked biofilms. In short, the presence of amide crosslinkages gives a better improvement in tensile properties of CS/EFB biofilms.

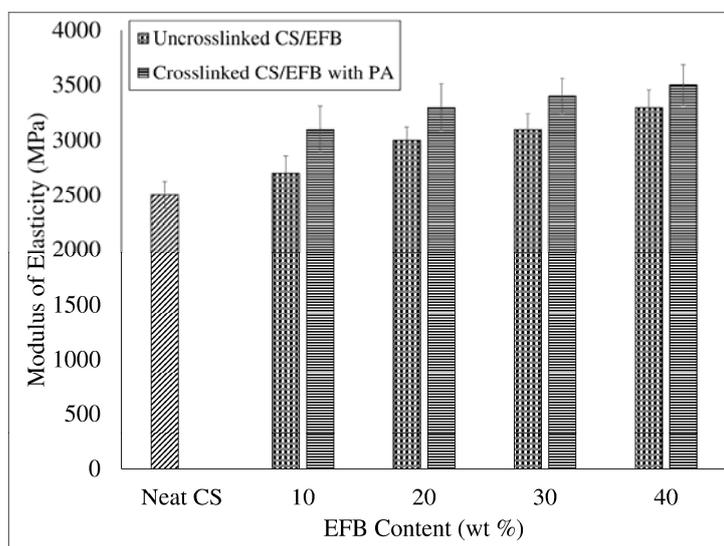


Figure 3: Modulus of elasticity of uncrosslinked and crosslinked CS/EFB biofilms with PA.

3.2 FTIR Analysis

Figure 4 presents the FTIR spectra of uncrosslinked and crosslinked CS/EFB biofilms with PA. Table 2 summarises the major functional groups of uncrosslinked and crosslinked CS/EFB biofilms with PA. From Figure 4, it can be observed that the band at 3323 cm^{-1} and 2910 cm^{-1} correspond to $-\text{OH}$ and $-\text{CH}$ groups, respectively, whereas the peaks at 1644 cm^{-1} and 1550 cm^{-1} correspond to amide I and amide II. Moreover, the peak at 1407 cm^{-1} is attributed to $-\text{CH}_2$ deformation from cellulose or $-\text{CH}$ group deformation from lignin. The C–N amino and C–H group deformations were attributed by 1327 cm^{-1} and 1265 cm^{-1} . The peaks of C–O–C and C–O groups were in the range of 1000 cm^{-1} to 1150 cm^{-1} . However, the spectrum of the crosslinked CS/EFB biofilm with PA shows lower intensity peak at 3343 cm^{-1} compared to the uncrosslinked biofilm. This indicates that the inclusion of the PA crosslinking agent reduced the hydrophilicity of CS/EFB biofilms.

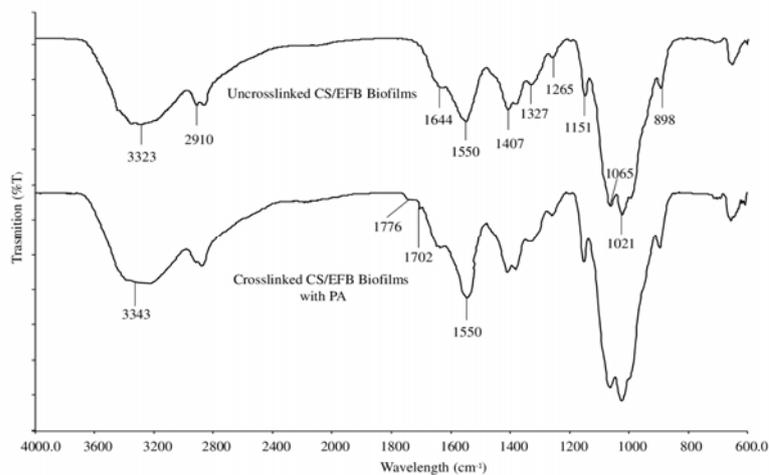


Figure 4: FTIR analysis of uncrosslinked and crosslinked CS/EFB biofilms with PA.

Table 2: The major functional groups of uncrosslinked and crosslinked CS/EFB biofilms with PA.

Biofilm	Frequency (cm ⁻¹)	Functional groups	Intensity (%)
Uncrosslinked CS/EFB biofilm	3323	-OH stretching	
	2910	-CH stretching	
	1644	Amide I	
	1550	Amide II	
	1407	-CH ₂ bond	-
	1327	C-N stretching (amino group)	
	1265	-CH stretching	
Crosslinked CS/EFB biofilm with PA	1000-1150	C-O-C, C-O bonds	
	3343	-OH stretching	Decreased (3.2)
	1776	C=O stretching (from anhydride)	New peak
	1702	C=O stretching (carboxylic groups)	New peak
	1550	C-N (amide bond)	Increased (2.0)

Two new peaks at 1702 cm⁻¹ and 1776 cm⁻¹ appeared, which are ascribed to the carboxylic groups (hydrolysis products of intermediated imide) and C=O groups from anhydride, respectively. Besides, the intensity peak at 1550 cm⁻¹ increased due to the presence of amide crosslinkages between CS and PA overlapped with

free primary amines. Similar finding was reported by Tangpasuthadol et al.²³, who studied the surface modification of CS films by anhydride crosslinker to improve the properties of CS films. The FTIR results show a new absorption peak at 1710 cm^{-1} and 1770 cm^{-1} attributed to the C=O stretching vibration of carboxyl groups. The proposed schematic crosslinking reaction between CS and PA is illustrated in Figure 5.

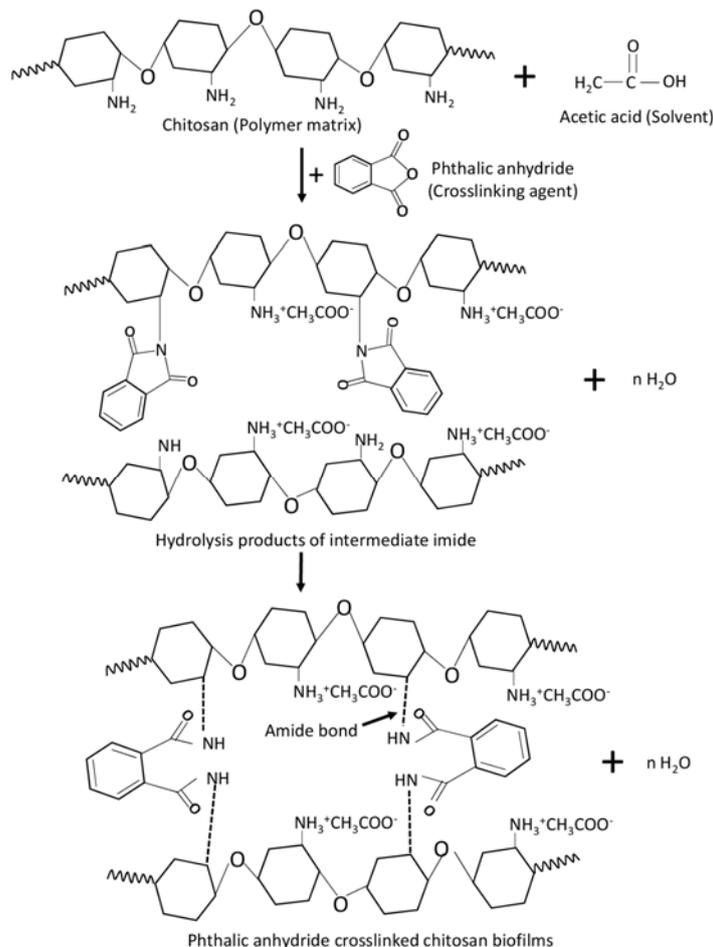


Figure 5: Proposed schematic crosslinking reaction between chitosan and phthalic anhydride.

3.3 Gel Fraction

In general, the gel fraction determination can be investigated the presence of crosslinkages in polymer.²⁴ The gel fraction of uncrosslinked and crosslinked

CS/EFB biofilms with PA is tabulated in Table 3. The gel fraction of CS/EFB biofilms increased with the increasing EFB content from 10 wt% to 40 wt% because the EFB natural filler would not dissolved in acetic acid. Nevertheless, the neat CS film was partially dissolved in acetic acid during swelling process as the CS formed a semi crystalline structure in solid state. This crystal structure hardly dissolves in acetic solution.²⁵ On the other hand, the crosslinked CS/EFB biofilms with PA exhibited higher gel fraction in comparison with uncrosslinked biofilms. This is due to the presence of amide linkages in biofilms.

Table 3: Gel fraction of uncrosslinked and crosslinked CS/EFB biofilms.

Biofilms	Gel fraction (%)
Neat CS	46.12
Uncrosslinked CS/EFB (80:20)	52.14
Uncrosslinked CS/EFB (60:40)	58.41
Crosslinked CS/EFB (80:20)	72.56
Crosslinked CS/EFB (60:40)	76.87

3.4 Differential Scanning Calorimetry (DSC)

The DSC curve of uncrosslinked and crosslinked CS/EFB biofilms is illustrated in Figure 6. The DSC data of CS/EFB biofilms is summarised in Table 4. CS biofilms easily absorbed moisture. Thus, to eliminate the moisturising effect, two cycles of heating and cooling were introduced. Similar finding was reported by a few studies.^{26,27} The glass transition temperature (T_g) of neat CS film occurred at approximately 169°C, as shown in Table 4. As the EFB content increased, the T_g shifted to a higher temperature due to the reduction of plasticity of the CS matrix. The presence of EFB restricted the chain mobility resulting in a higher T_g of CS/EFB biofilms. Nevertheless, the crosslinked CS/EFB biofilms with PA at 20 wt% and 40 wt% of EFB content showed higher T_g than uncrosslinked CS/EFB biofilms. This was due to the presence of new inters amide linkages, which become rigid and resist deforming. This observation is in line with modulus of elasticity results which was explained earlier.

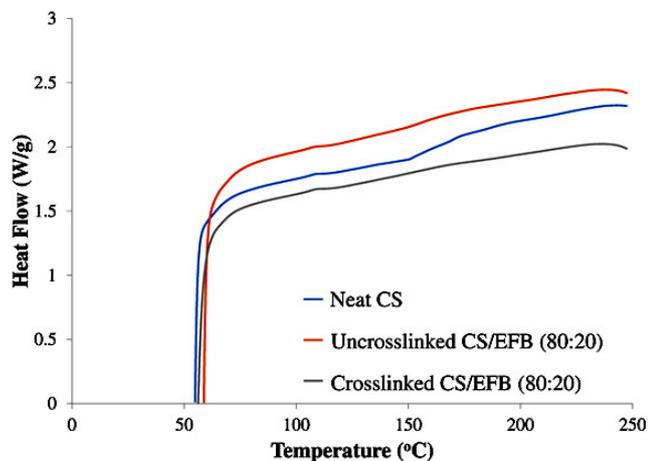


Figure 6: DSC curve of neat CS film, uncrosslinked and crosslinked CS/EFB biofilms with PA.

Table 4: DSC data of CS/EFB uncrosslinked and crosslinked biofilms.

Biofilms	T_g (°C)
CS	169
Uncrosslinked CS/EFB (80/20)	156
Uncrosslinked CS/EFB (60/40)	165
Crosslinked CS/EFB (80/20)	164
Crosslinked CS/EFB (60/40)	185

3.5 Morphology Study

Figure 7 shows the SEM micrograph of homogenous surface and matrix tearing of neat CS film. Figures 8 (a) and (b) illustrate the SEM micrograph of tensile fractured surface for uncrosslinked and crosslinked CS/EFB biofilms at 40 wt% EFB content, respectively. Figure 8(a) shows the rough surface with the addition high content of EFB. Additionally, the micrograph from Figure 8(a) exhibits some detachment of EFB from the CS, agglomeration and poor dispersion. This indicates poor interaction between EFB and CS matrix. This result is in line with the finding that the tensile strength of biocomposite films decreased with increasing EFB content, as discussed earlier. The SEM micrograph of crosslinked biofilm as demonstrated in Figure 8(b) shows less EFB filler detachment, less agglomeration, some embedding of EFB filler, and better dispersion in CS matrix. It can be seen that the surface is smoother as compared to without the addition of

phthalic anhydride. This indicates that the presence of phthalic anhydride enhanced the filler-matrix interfacial adhesion and filler dispersion.

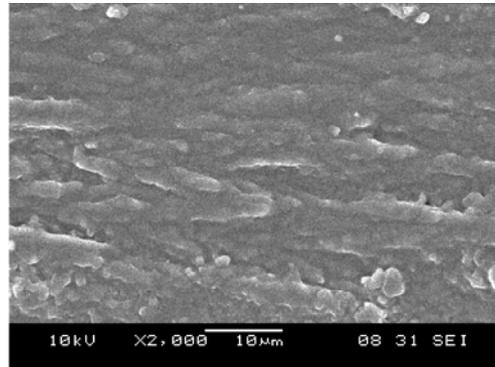


Figure 7: SEM micrograph of neat CS film.

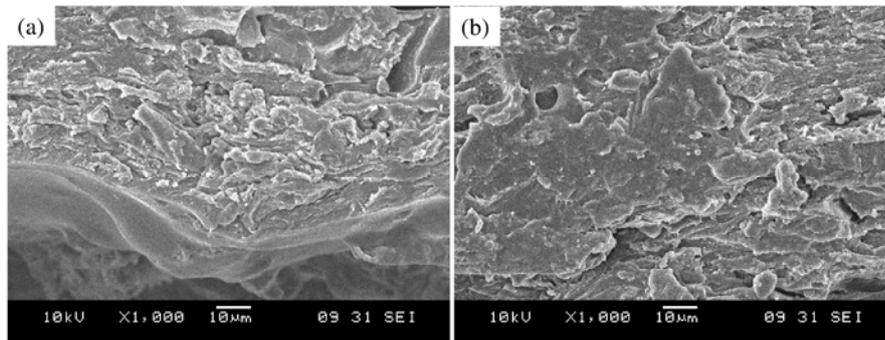


Figure 8: SEM micrograph of (a) uncrosslinked CS/EFB biofilm; and (b) crosslinked CS/ EFB biofilm at 40 wt% of EFB content, respectively.

4. CONCLUSION

In summary, changes in EFB content and chemical modification with PA had effects on tensile and thermal properties of EFB/CS biofilms. It has been found that the tensile strength and elongation at break decreased, but modulus of elasticity increased with the increasing EFB content. However, the PA improved the tensile properties of EFB/CS biofilms due to formation of amide crosslinkages and better filler-matrix adhesion. The T_g of biocomposite films shifted to a higher temperature with the increasing amount of EFB. After chemical modification with PA, a higher T_g was observed as compared to neat CS film and uncrosslinked EFB/CS biofilms. The gel fraction of uncrosslinked and crosslinked CS/EFB biofilms increased with the increasing EFB content.

Crosslinked CS/EFB biofilms exhibited higher gel fraction than uncrosslinked biofilms due to the presence of amide bonds in biofilms.

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